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# Diagnostic value of meat juice in early detection of *Classical swine fever virus* infection

Louise Lohse,<sup>1</sup> Åse Uttenthal, Thomas Bruun Rasmussen, Jens Nielsen

**Abstract.** To evaluate the diagnostic potential of meat juice for early detection of *Classical swine fever virus* (CSFV), meat juice and serum samples from pigs experimentally infected with different strains of CSFV were compared for virus load. From all samples, viral RNA was extracted by automated procedure before real-time reverse transcription polymerase chain reaction analysis was performed. Viral RNA was detected in meat juice, but at a lower level than in corresponding serum. Sensitivity was calculated to 91% and specificity to 97%. Disagreements between meat juice and serum results were found when samples originated from pigs infected with low virulence CSFV strains and/or when samples were collected within the first days after infection. In conclusion, while not the first choice for sample material for CSFV diagnosis, meat juice may constitute a useful alternative for herd-based studies or when blood and/or target organ material is not available. Strain virulence and time points for sample collection after infection are factors of importance for diagnostic success.

**Key words:** *Classical swine fever virus*; meat juice; reverse transcription polymerase chain reaction; swine.

Classical swine fever (CSF) is a serious, highly contagious disease in swine. The causative agent is *Classical swine fever virus* (CSFV), which is a positive-stranded RNA virus belonging to the *Pestivirus* genus in the *Flaviviridae* family.<sup>10</sup> Although many countries in Europe are free from the disease, sporadic or massive new outbreaks are still occasionally detected.<sup>3,4,7,11</sup> Clinical signs of CSF are not pathognomonic, as confirmation of this disease is usually based on laboratory diagnosis. Thus, the availability of blood and/or tissue material is essential. Tissue samples from target organs of CSFV, including tonsils, spleen, kidney, ileum, and lymph nodes, constitute the optimal choice for virus detection (2002, Classical swine fever diagnostic manual, document number C 381, eur-lex.europa.eu 32002D0106 EN; OIE: 2010, Classical swine fever (hog cholera), www.oie.int/eng/normes/mmanual/a\_summry.htm). In situations under less optimal conditions, such sample material may not be available. Therefore, the use of alternative materials may be justified.

Skeletal muscle tissue can be easily collected for laboratory examination from abattoir carcasses, wild boars killed in road accidents, and imported meat products. From this material, meat juice can be extracted for further diagnostic analysis. Previous studies have performed pathogen and/or antibody analyses of meat juice for laboratory detection of several infections including salmonellosis, Aujeszky disease, porcine reproductive and respiratory syndrome, trichinellosis,<sup>1,6,8,9</sup> and CSF.<sup>5,14</sup> However, the existence of numerous CSFV strains of different virulence make the diagnostic

approach for CSFV infection less straightforward, as virus distribution may differ with virulence.<sup>2</sup> This fact could influence the suitability of meat juice as a potential diagnostic sample material.

In the present study, the diagnostic potential of meat juice for detection of CSFV RNA was tested by examination of skeletal muscle tissue samples from pigs experimentally infected with 7 CSFV strains of different genotypes and virulence. The performance of meat juice was compared with contemporary analyses of serum collected from the same animal at the same time points. All material originated from CSFV infection experiments in pigs performed at the National Veterinary Institute (Lindholm, Kalvehave, Denmark) in accordance with the requirements of the Danish Animal Experiments Inspectorate.

For evaluation, a real-time reverse transcription polymerase chain reaction (RT-PCR) assay method in an automated RNA extraction system was used on sampled material from a total of 85 pigs. All samples were collected from pigs at time of euthanasia, which varied from day post-inoculation (dpi) 5–28. For pigs infected with CSFV-Bergen, CSFV-Glentorf, CSFV-Romania, CSFV-Israel, and mock-inoculated control pigs, the experimental design included sequential

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**Table 1.** Diagnostic value of meat juice in early detection of classical swine fever infection: virulence and genotype information of the 7 Classical swine fever virus (CSFV) strains used and results after real-time reverse transcription polymerase chain reaction (RT-PCR) examination of serum and meat juice samples.\*

Virulence designation	Strain	Strain	Genotype	No. of pigs examined	dpi	Sample material			
						Meat juice		Serum	
						RT-PCR positive	RT-PCR negative	RT-PCR positive	RT-PCR negative
Low	CSFV-Glentorf	CSF0911	1.1	10	5–21	2	8	3	7
	CSFV-Bergen	CSF0906	2.2	10	5–21	4	6	5	5
Moderate	CSFV-Paderborn	CSF0277	2.1	13	15–24	13	0	13	0
	CSFV-Eystrup	CSF0910	1.1	10	8–28	5	5	5	5
High	CSFV-Romania	CSF1019	2.3	10	5–18	8	2	10	0
	CSFV-Kozlov	CSF0382	1.1	9	6–8	9	0	9	0
	CSFV-Israel	CSF1047	2.1	10	5–11	10	0	10	0
Control	Mock inoculated	NR	NR	13	5–28	0	13	0	13
Total				85		51	34	55	30

\*dpi = day post-infection; NR = not relevant. Strain ID and genotype designation is obtained from the CSFV database from The European Central Reference Laboratory. RT-PCR-positive and -negative results indicate threshold cycle <40 and ≥40, respectively.

killings in groups of 3–4 on dpi 5, 10, and 21/22, respectively, except for individual pigs, which had to be euthanized at an earlier time point for animal welfare reasons. After collection, the sampled materials were prepared for further examination as follows. Serum was obtained by centrifugation of blood collected in plain tubes with no anticoagulant at 237 g for 10 min and then stored at –40°C until use. Meat juice was obtained from samples of the quadriceps muscle. Thus, muscle tissue was cut into pieces of approximately 2 cm × 2 cm × 2 cm, put into plastic bags, frozen at –20°C, and followed by gentle thawing at 5°C. Through this process, the released meat juice was collected without further preparation into cryotubes and stored at –40°C until use. For each analysis, 100 µl of sample material (serum or meat juice) was used.

The RNA was extracted with a commercial nucleic acid isolation kit,<sup>a</sup> according to the manufacturer's instructions using an automated robot.<sup>a</sup> After RNA extraction, the material was stored at –80°C in the sample cartridge until PCR was carried out. Real time RT-PCR was performed as previously described<sup>12</sup> using the primers CSFV6 and CSFV7 together with a CSFV-specific TaqMan probe (5'-FAM-CCCTGG GTGGTCTAAGTCCTGAGTACAG-TAMRA-3'). The reactions were cycled,<sup>b</sup> and, to the fluorescence data obtained, a threshold cycle (Ct) value was assigned to each sample as a measure for the virus quantity. The relative quantity of viral RNA (the viral load) in each sample was expressed directly as Ct values without recalculating the genome equivalents.

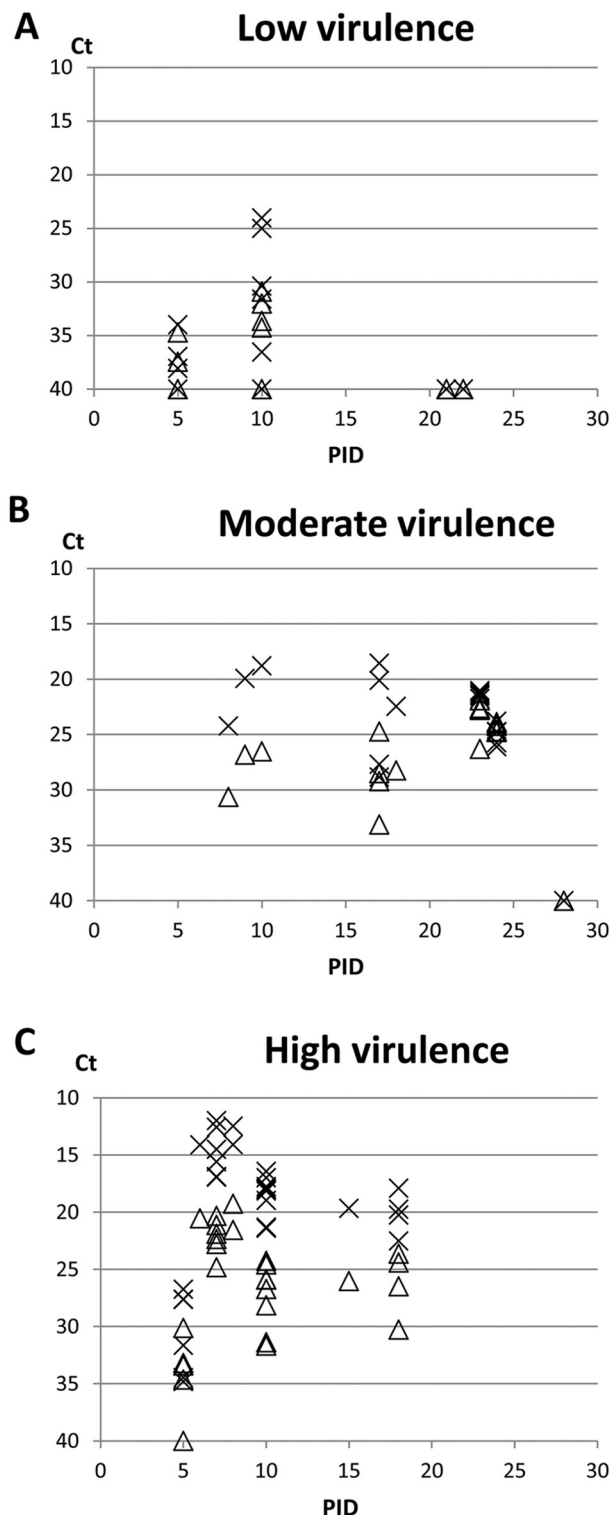
Data obtained from the serum and meat juice analyses of pigs inoculated with the different CSFV strains were compared. Based on clinical records from the performed experiments, data sets from individual strains were designated into low, moderate, or high virulence (Table 1). To study the possible association between serum and meat juice, a Spearman rank correlation was calculated on the total data set of

inoculated animals, and the correlation coefficient (rs) was determined.

The Ct values of serum and meat juice from individual pigs were depicted with relation to infection day in the experiment and illustrated in Figure 1A–C. The results indicate that CSFV RNA is detected in meat juice, yet in a lower quantity than in serum. Furthermore, throughout the entire experimental period, negative samples of meat juice as well as serum will occur in a group of pigs infected with a CSFV strain of low virulence (Fig. 1A).

All sample results were assigned to either positive (Ct value < 40) or negative (Ct value ≥ 40) value (Table 1), and by comparison, meat juice sensitivity was calculated to 91% and specificity to 97% of serum. The coefficient of concordance for the 2 tested materials was 93%, indicating that a strong correlation exists. Six pairs of samples were in disagreement; in 5 cases, serum tested positive for CSFV RNA while negative in meat juice, and in 1 case, the relationship was reversed. Four of the disagreements were related to strains in the category of low virulence: 1 sample from a CSFV-Glentorf-infected pig (dpi 5) and 3 samples from CSFV-Bergen-infected pigs (dpi 5/10). Two disagreements were related to the category of high virulence; both samples were collected from CSFV-Romania-infected pigs (dpi 5). All 13 mock-infected control pigs tested negative in serum as well as in meat juice. The calculated correlation coefficient (rs) was = 0.89, with a 95% confidence interval of (0.82; 0.93) and a *P* value < 0.0001, which indicates that there is quite a strong positive correlation between the 2 sample materials.

In order to address situations where blood and/or target organ material for CSFV diagnosis is not available, the potential of meat juice as diagnostic substrate was evaluated. Previous studies on meat juice and muscular tissue reflect different results and diverging opinions regarding the diagnostic



**Figure 1.** Threshold cycle (Ct) values obtained from examined sample material after real-time reverse transcription polymerase chain reaction procedure. Depicted Ct values are shown for individual pigs and related to day post-infection (PID). Results from the individual groups of pigs inoculated with different *Classical swine fever virus* strains are designated into categories of virulence (A–C), based on clinical records. Δ = meat juice; × = serum.

value of this material.<sup>5,13,14</sup> Based on the present study with CSFV strains of different virulence, the levels of CSFV RNA in meat juice and corresponding serum samples were compared. By comparison of data from all inoculated pigs, an overall sensitivity of 91% for detection of CSFV in meat juice was calculated. The results obtained in the current study support previous work,<sup>5</sup> which concluded that meat juice is most suitable as substrate for diagnosis of CSFV from pigs infected with strains of high or moderate virulence. In low virulence CSFV infections, virus detection in meat juice as well as in other nonlymphatic organs can be problematic due to low virus load and sparse tissue distribution in the infected pig. Therefore, for low virulent strains, demonstration of serum antibodies several weeks after infection may be of diagnostic value. Furthermore, the results suggest that time point after infection has an important effect in sample collection, as some virus strains, even when classified as highly virulent, will not be disseminated to non-virus-targeted organs within the body of the pig until the end of the first week after infection. Such a problem was encountered in pigs inoculated with CSFV-Romania, where demonstration of viral RNA in meat juice only was possible in 1 out of 3 pigs at dpi 5. Corresponding serum samples were all positive for viral RNA, indicating ongoing viremia. The finding does not correspond with conclusions from a Swiss study,<sup>13</sup> which could not demonstrate any influence of time point of infection for sample collection for detection of CSFV RNA. However, the results from the Swiss and the present study are not directly comparable as the presented data are based on homogenized muscle tissue and meat juice, respectively. In a previous Danish study,<sup>14</sup> a comparison of virus detection in meat juice and corresponding homogenized quadriceps muscle tissue revealed lower Ct values in meat juice than in corresponding muscle tissue samples, indicating that a higher RT-PCR sensitivity for the former sample material exists. The finding is consistent with the Swiss study<sup>13</sup> that could not recommend muscular tissue as diagnostic material for CSFV, regardless of strain virulence, and found that only 58% of the tested samples were CSFV positive by RT-PCR compared with parallel examination of target organs. It is speculated whether the lower sensitivity of muscle tissue samples compared with meat juice might be due to PCR inhibiting factors released by tissue homogenization or dilution of virus content in the muscle tissue preparation process.

In conclusion, the results from the present study indicate that the use of meat juice as diagnostic material for CSFV infection is justified under certain conditions (e.g., when used as material for herd-based screening). However, meat juice cannot be recommended as an isolated diagnostic material for early detection of CSFV infection in individual pigs when blood and/or target organ material is available, as the diagnostic sensitivity for meat juice is lower than that for serum. Potential use of meat juice as a reliable diagnostic sample will depend on the actual strain virulence and on the relative time of sample collection in the specific disease situation.

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### Sources and manufacturers

- a. MagNA Pure LC Total Nucleic Acid Isolation kit, Roche A/S, Hvidovre, Denmark.
- b. MX4000, Stratagene, AH-diagnostics, Aarhus, Denmark.

### Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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### References

1. Beck R, Gaspar A, Mihaljević Z, et al.: 2005, Evaluation of ELISA for detection of *Trichinella* antibodies in meat juice samples of naturally infected pigs. *Vet Parasitol* 132:91–95.
2. Belák K, Koenen F, Vanderhallen H, et al.: 2008, Comparative studies on the pathogenicity and tissue distribution of three virulence variants of classical swine fever virus, two field isolates and one vaccine strain, with special regard to immunohistochemical investigations. *Acta Vet Scand* 50:34.
3. Elber AR, Stegeman A, Moser H, et al.: 1999, The classical swine fever epidemic 1997–1998 in The Netherlands: descriptive epidemiology. *Prev Vet Med* 42:157–184.
4. Fritzemeier J, Teuffert J, Greiser-Wilke I, et al.: 2000, Epidemiology of classical swine fever in Germany in the 1990s. *Vet Microbiol* 77:29–41.
5. Kaden V, Lange E, Nagel-Kohl U, Bruer W: 2009, Meat juice as diagnostic sample for virological and serological diagnosis of classical swine fever. *Dtsch Tierarztl Wochenschr* 116:173–179.
6. Le Potier MF, Fournier A, Houdayer C, et al.: 1998, Use of muscle exudates for detection of anti-gE antibodies to Aujeszky's disease virus. *Vet Rec* 143:385–387.
7. Moennig V, Floegel-Niesmann G, Greiser-Wilke I: 2003, Clinical signs and epidemiology of classical swine fever: a review of new knowledge. *Vet J* 165:11–20.
8. Mortensen S, Strandbygaard B, Bøtner A, et al.: 2001, Monitoring porcine reproductive and respiratory syndrome virus infection status in swine herds based on analysis of antibodies in meat juice. *Vet Res* 32:441–453.
9. Nielsen B, Ekeröth L, Bager F, Lind P: 1998, Use of muscle fluid as a source of antibodies for serologic detection of *Salmonella* infection in slaughter pig herds. *J Vet Diagn Invest* 10:158–163.
10. Paton DJ, McGoldrick A, Greiser-Wilke I, et al.: 2000, Genetic typing of classical swine fever virus. *Vet Microbiol* 73:137–157.
11. Pol F, Rossi S, Mesplède A, et al.: 2008, Two outbreaks of classical swine fever in wild boar in France. *Vet Rec* 162:811–816.
12. Rasmussen TB, Uttenthal Å, Reimann I, et al.: 2007, Virulence, immunogenicity and vaccine properties of a novel chimeric pestivirus. *J Gen Virol* 88:481–486.
13. Thür B, Hofmann MA: 1998, Comparative detection of classical swine fever virus in striated muscle from experimentally infected pigs by reverse transcription polymerase chain reaction, cell culture isolation and immunohistochemistry. *J Virol Methods* 74:47–56.
14. Uttenthal A, Storgaard T, Oleksiewicz MB, de Stricker K: 2003, Experimental infection with the Paderborn isolate of classical swine fever virus in 10-week-old pigs: determination of viral replication kinetics by quantitative RT-PCR, virus isolation and antigen ELISA. *Vet Microbiol* 92:197–212.